Cell Culture Protocol for Raji cells

Raji (ATCC number CCL-86): Burkitt's Lymphoma; B lymphocyte cell line
Growth medium: RPMI Mediujm 1640 (GIBCO # 21870) + 10% FBS + 2mM L-Glutamine + 100 units/ml penicillin + 100 micro-g/ml streptomycin (GIBCO # 15140-122).

Protocol for Thawing Raji Cells:
1. Take out the Raji stock vial from liquid nitrogen (we freeze at 1x10^6 cells per vial) and thaw it in 37° water bath. Suspend thawed cells in 5 ml growth media.
2. Centrifuge at 1000 rpm for 3 min, discard media.
3. Resuspend cell pellet in 15ml growth media and transfer cells into a 75 sq. cm. tissue culture flask. Cells are grown in a 37ºC incubator at 5% CO2.

Protocol for Subculturing of Raji Cells:
Change medium every 2 to 3 days, and split cultures when they reach 2-3x10^6 viable cells/ml.

1. Transfer cell suspension (in growth medium) to centrifuge tubes and spin at 1000rpm for 3minutes.
2. Aspirate supernatant and add 10mls of fresh growth media
3. Count cells, and transfer 0.5-5X10^5 viable cells/ml to a new culture vessel (this can either be a spinner flask or a tissue culture flask that's placed horizontally a 37ºC incubator at 5% CO2.)

Note: We typically split cells at a ratio of 1:10 every 2 to 3 days using the abovementioned seeding conditions (and we grow a back-up maintenance plate, in addition to the culturing vessel used for experiments, by adding about 1x10^5 cells per 75 sq. cm flask).

Freezing of Raji Cells:
Cells can be stored as a stock in liquid nitrogen at 1x10^6 cells/ml in growth medium containing 10% DMSO.